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(54) Title: A NEW ENZYME ISOLATED FROM A *BIFIDOBACTERIUM*

(57) Abstract: The present invention concerns a new  $\beta$ -galactosidase with transgalactosylating activity isolated from *Bifidobacterium bifidum* and a truncated enzyme where the C-terminal end of the  $\beta$ -galactosidase protein has been deleted resulting in an enzyme with a higher transgalactosylating activity than hydrolase activity. When lactose is used as a substrate, galacto-oligosaccharides are products of the transgalactosylase activity. Galacto-oligosaccharides enhance growth of health-promoting *Bifidobacterium* that may be used in a number of applications in the dairy industry.



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## A new enzyme isolated from a *Bifidobacterium*

### Technical field of invention

5 The present invention concerns improvement of fermented  
diary products. In particular, the invention concerns a  
 $\beta$ -galactosidase with transgalactosylating activity. More  
particular the inventions concerns a  $\beta$ -galactosidase  
isolated from *Bifidobacterium bifidum* where the C-  
10 terminal end of the protein has been deleted and the  
resulting truncated enzyme has higher  
transgalactosylating activity than hydrolase activity.  
When lactose is used as a substrate, galacto-  
oligosaccharides are products of the transgalactosylase  
15 activity. Galacto-oligosaccharides enhance growth of  
health-promoting *Bifidobacterium* that may be used in a  
number of applications in the dairy industry.

### Background of the invention

20 The genus *Bifidobacterium* is one of the most commonly  
used types of bacteria cultures in the dairy industry for  
fermenting a variety of dairy products. Ingestion of  
*Bifidobacterium*-containing products furthermore has a  
25 health-promoting effect. This effect is not only achieved  
by a lowered pH of the intestinal contents but also by  
the ability of *Bifidobacterium* to repopulate the  
intestinal flora in individuals who have had their  
intestinal flora disturbed by for example intake of  
30 antibiotics. *Bifidobacterium* furthermore has the  
potential of outcompeting potential harmful intestinal  
micro-organisms.

Galacto-oligosaccharides are known to enhance the growth of *Bifidobacterium*. This effect is likely achieved through the unique ability of *Bifidobacterium* to exploit galacto-oligosaccharides as a carbon source. Dietary  
5 supplement of galacto-oligosaccharides is furthermore thought to have a number of long-term disease protecting effects. For example, galacto-oligosaccharide intake has been shown to be highly protective against development of colorectal cancer in rats (Wijnands, et al., 1999). There  
10 is therefore a great interest in developing cheap and efficient methods for producing galacto-oligosaccharides for use in the industry for improving dietary supplements and dairy products.

15 The enzyme  $\beta$ -galactosidase (EC 3.2.1.23) usually hydrolyses lactose to the monosaccharides D-glucose and D-galactose. In the normal enzyme reaction of  $\beta$ -galactosidases, the enzyme hydrolyses lactose and transiently binds the galactose monosaccharide in a  
20 galactose-enzyme complex that transfers galactose to the hydroxyl group of water, resulting in the liberation of D-galactose and D-glucose. However, at high lactose concentrations some  $\beta$ -galactosidases are able to transfer galactose to the hydroxyl groups of D-galactose  
25 or D-glucose in a process called transgalactylation whereby galacto-oligosaccharides are produced.

Enzymes capable of transgalactosylation have been isolated from a wide range of micro-organisms, including  
30 bacteria and yeasts. The observation that galacto-oligosaccharides enhance the growth of health-promoting *Bifidobacterium* has stimulated investigations of *Bifidobacterium* and their  $\beta$ -galactosidase enzymes. Two DNA sequences of *B. breve* and *B. longum*  $\beta$ -galactosidase

genes have been deposited in GeneBank (accession numbers E5040 and AJ242596, respectively). Dumortier et al. (1994) have reported that *B. bifidum* DSM 20215 contains three  $\beta$ -galactosidases and one of these enzymes has  
5 trans-galactosylating properties. However, no identification of the enzyme possessing this activity or any sequence of the enzyme or the corresponding gene from *B. bifidum* DSM 20215 has been published.

10 Production of galacto-oligosaccharides by the use of  $\beta$ -galactosidases has been reported in several papers. For example,  $\beta$ -galactosidase from *E. coli* has been shown to produce oligosaccharides at high lactose concentrations (0.5 M or approximately 20% lactose; Huber et al. 1976).  
15 Various thermophilic microorganisms have been shown to produce oligosaccharides at high temperatures and high lactose concentrations, e.g. *Sterigmatomyces elviae* can produce 39% oligosaccharides from 20% lactose at 60°C (Onishi & Tanaka, 1995), and *Saccharopolyspora*  
20 *rectivirgula* can synthesize 41% oligosaccharides in 1.75 M lactose at 70°C (Nako et al., 1994).

However, the enzymes described above all have the drawbacks of requiring either high temperatures or high  
25 lactose concentrations or both in order to exhibit significant transgalactosylase activity. There is thus a need for developing cheaper and more efficient methods of producing galacto-oligosaccharides for use in the industry.

30

#### Summary of the invention

The present invention describes a new  $\beta$ -galactosidase from *Bifidobacterium bifidum*. A truncated version of the

enzyme has surprisingly been shown to have a high transgalactosylating activity. When the truncated enzyme, or a host cell expressing the recombinant truncated enzyme is incubated with lactose under appropriate  
5 conditions, galacto-oligosaccharides are produced at a high efficiency. Presence of galacto-oligosaccharides in dairy products or other comestible products have the advantage of enhancing the growth of health-promoting *Bifidobacterium* in the product or in the intestinal flora  
10 of the consumer after intake of the product or both.

#### Brief description of the drawings

##### Figure 1:

15 OLGA5 sequence. DNA and protein sequence of the OLGA5  $\beta$ -galactosidase from *Bifidumbacterium bifidum*. The signal sequence is shown in bold and the part of OLGA5 gene deleted in OLGA347 is shown in italics. The *Bgl*III site used to create the deletion is highlighted.

20

##### Figure 2:

Comparison of  $\beta$ -galactosidase active site regions. Alignment of regions around the catalytic Glu461 residue (highlighted) from *E. coli*. The sequences are identified  
25 by their database accession numbers. 6-phospho- $\beta$ -galactosidase sequences are marked with a (P).

##### Figure 3:

Neighbour joining analysis of the alignment in Figure 1, where the *Sulfolobus* sequences were used as an outgroup.  
30 Results from a bootstrap analysis (n = 100) are shown for the junctions with a value above 80.

##### Figure 4:

OLGA5 transgalactosylase activity. Total cell lysate of *E. coli* cells harbouring the OLGA5 gene in a plasmid were incubated with 0.4 M lactose at 37°C for 20 hours. A 50 µl total reaction volume contained the indicated amounts of total cell lysate. Reaction samples were analysed on a silica gel TLC plate. The plate was sprayed with Orcinol reagent to visualise the sugars.

Figure 5:

C-terminal deletions of OLGA5 β-galactosidase. A 1752 amino acid open reading frame encodes the OLGA5 β-galactosidase, where the starting 32 amino acids likely represent a signal peptide (white box). Deletion mutants of OLGA5 were constructed using the indicated restriction sites. Lysates prepared from bacterial cultures grown over night were used for measurement of β-galactosidase activity, and the relative results are shown to the right of the respective constructs. Restriction enzyme symbols used: *Bgl*III (B), *Eco*RI (E), *Eco*RV (V), *Hind*III (H), *Kpn*I (K), *Nru*I (N), *Pst*I (P).

Figure 6:

TLC analysis of transgalactosylase activity. Total cell lysates for the two tested deletion mutants, OLGA347 and OLGA345, were used in the indicated amounts to react with 0.4 M lactose in 50 µl total volume. The reactions were incubated at 37°C for 20 hours. Samples were analysed on a silica gel TLC plate. The plate was sprayed with Orcinol reagent to visualise the sugars.

Figure 7:

Oligosaccharides produced by OLGA347. The indicated amounts of OLGA347 total cell lysate were incubated with 15% lactose in a total volume of µl for 21 hours at 37°C.



Radioactive lactose that was labelled with  $^{14}\text{C}$  in the glucose C-1 position was used. Samples were separated on a TLC plate and quantitated by use of a phospho-imager. A: Image used for measurement of  $^{14}\text{C}$ -signals from  
5 lactose, glucose and galacto-oligosaccharides (GOS) spots. B: Measured  $^{14}\text{C}$ -signals after subtraction of background (blind lane).

Figure 8:

10 HPLC measurement of OLGA347 enzyme reaction products. Reactions in 10%, 20% and 40% lactose were performed using the indicated amounts of OLGA347 total cell lysate. A total volume of 200  $\mu\text{l}$  was used and the reactions were  
15 subjected to HPLC analysis and standard curves were used to convert the observed peak areas to concentrations (mg/ml). A: Obtained mg/ml saccharide after OLGA347 reaction with 10% lactose. B: Obtained mg/ml saccharide after OLGA347 reaction with 20% lactose. C: Obtained  
20 mg/ml saccharide after OLGA347 reaction with 40% lactose. D: Plot of results from the 10% reaction. The resulting amount of galacto-oligosaccharides is calculated as the amount of lactose not recovered as glucose or galactose ("GOS").

25

**Detailed description of the invention**

The first aspect of the invention concerns a new  $\beta$ -galactosidase, OLGA5 (SEQ ID NO:1 and SEQ ID NO:2), from  
30 *Bifidobacterium bifidum* that has been isolated and characterised. *E. coli* cells were transformed with a plasmid containing insertions consisting of *Pst*I digested chromosomal DNA from *B. bifidum*. Clones with  $\beta$ -galactosidase activity were selected on plates containing

a chromogenic  $\beta$ -galactosidase substrate. One of the positive colonies contained a plasmid with an insert of approximately 20 kb, pOLGA5 (SEQ ID NO:1). Sequencing of the DNA sequence revealed that the deduced amino acid sequence of OLGA5  $\beta$ -galactosidase (SEQ ID NO:2) is approximately twice as long as the presently known  $\beta$ -galactosidases and it furthermore shows a surprisingly low degree of sequence homology with known  $\beta$ -galactosidases. Expression of recombinant OLGA5 in *E. coli* revealed that the enzyme, in addition to lactose hydrolysing activity, also exhibited transgalactosylating activity. The C-terminal part of the OLGA5 enzyme showed no homology to known  $\beta$ -galactosidases. A variety of OLGA5 C-terminal deletion mutants were subsequently constructed and the resulting enzymes were investigated for their hydrolytic and transgalactosylating activity.

A second aspect of the invention concerns deletion mutants of OLGA5, e.g. OLGA347. Out of several C-terminal deletion mutants, OLGA347 which has a 578 amino acid C-terminal deletion, showed the most pronounced increased level of oligosaccharides produced when incubated with lactose even at relatively low lactose concentrations. The enzyme apparently transferred virtually all galactose molecules onto galactose or glucose. Deletion of the C-terminal end of OLGA5 hence converted the enzyme from a hydrolytic OLGA5  $\beta$ -galactosidase to a transgalactosylating OLGA347-transgalactosidase. Unlike other transgalactosylating  $\beta$ -galactosidases, including the native OLGA5 enzyme, the truncated  $\beta$ -galactosidase



OLGA347 transfers galactose onto acceptor sugar molecules at high frequency at all lactose concentrations examined.

In one embodiment, an expression vector with an insert  
5 encoding OLGA5, OLGA342, OLGA345, OLGA347, OLGA344, or  
any other OLGA5 variant is used. This expression vector  
can be transformed into a host cell selected from the  
group comprising *Bifidobacterium*, *Lactococcus*,  
*Lactobacillus*, *Streptococcus*, *Leuconostoc*, *Escherichia*,  
10 *Bacillus*, *Streptomyces*, *Saccharomyces*, *Kluyveromyces*,  
*Candida*, *Torula*, *Torulopsis* and *Aspergillus*. A cell of  
the genus *Bifidobacterium* is selected from the group  
consisting of *Bifidobacterium breve*, *Bifidobacterium*  
*longum*, *Bifidobacterium infantis*, *Bifidobacterium bifidum*  
15 and *Lactococcus lactis*. The cell is then cultured in a  
suitable culture medium under conditions permitting  
expression of for example an OLGA5 or an OLGA347 variant  
and the resulting enzyme is thereafter recovered from the  
culture.

20 In another embodiment of the invention, an OLGA5 variant  
is part of an expression vector, which can be transformed  
into any one of the above, mentioned host cells. The cell  
is then cultured in a suitable culture medium under  
25 conditions permitting expression of the OLGA5 variant and  
the resulting enzyme is thereafter recovered from the  
culture. The OLGA5 variant may contain any random  
mutation or any mutation generated by conventional  
molecular biology techniques. Any fragment of a mutated  
30 or a wild-type OLGA5 DNA molecule can be inserted into  
the expression vector. The fragment can be generated by  
PCR (polymerase chain reaction) or by means of any  
restriction sites present in the sequence or a  
combination of both. The procedures for generating OLGA5

variants are well known to a person skilled in the art. It is thus not critical to the present invention in which way the variant is obtained. The variants disclosed in the present text are obtained by subcloning by use of  
5 restriction sites present in the sequence.

Another aspect of the invention concerns use of one or more of the above mentioned cell types for producing a product selected from the group consisting of yoghurt,  
10 cheese, fermented dairy products, dietary supplements and probiotic comestible products. In this aspect, the technical effect of the enhanced growth of *Bifidobacterium* is used for improving the quality of the industrial products. Addition of galacto-oligosaccharides  
15 enhances the growth of health-promoting *Bifidobacterium*. Galacto-oligosaccharides produced by OLGA347 is thus much cheaper and easier to obtain compared to using native  $\beta$ -galactosidases for producing oligosaccharides.

20 Yet another aspect of the invention concerns the use of OLGA5, OLGA342, OLGA345, OLGA347, OLGA344 or any other OLGA5 variant or the use of any one or more of the above mentioned cell types for producing oligosaccharides. The  
25 oligosaccharides comprise, but are not limited to fructooligo-saccharides, galacto-oligosaccharides, isomalto-oligosaccharides, malto-oligosaccharides, lacto-sucrose and xylo-oligosaccharides.

30 In one embodiment of the invention, the oligosaccharides are produced by incubating the cell expressing the OLGA5 variant in a medium that comprises a disaccharide substrate such as for example lactulose, trehalose, rhamnose, maltose, sucrose, lactose, or cellobiose. The

incubation is carried out under conditions where oligosaccharides are produced. The cells may be part of a product selected from the group consisting of yoghurt, cheese, fermented milk products, dietary supplements, and probiotic comestible products. Alternatively, the oligo-saccharides can be recovered and subsequently be added to the product of interest before or after its preparation. Addition of oligosaccharides enhance growth of either *Bifidobacterium* alone or of *Bifidobacterium* in a mixed culture.

In another embodiment, the oligosaccharides are produced by incubating the OLGA5 variant in a medium that comprises a disaccharide substrate such as for example lactulose, trehalose, rhamnose, maltose, sucrose, lactose, or cellobiose. The incubation is carried out under conditions where oligosaccharides are produced. The medium comprising an OLGA5 variant and lactose may be part of a product selected from the group consisting of yoghurt, cheese, fermented milk products, dietary supplements, and probiotic comestible products. Alternatively, the oligo-saccharides can be recovered and subsequently be added to the product of interest before or after its preparation. Addition of oligosaccharides enhances growth of either *Bifidobacterium* alone or of *Bifidobacterium* in a mixed culture.

#### Definitions

" $\beta$ -galactosidase or a fragment thereof".  $\beta$ -galactosidase is defined as an enzyme capable of hydrolysing lactose to the monosaccharides D-glucose and D-galactose. A fragment of the  $\beta$ -galactosidase comprises 5-98%, preferably 40-95%

and most preferably 55-75% of the protein and the deletion preferably concerns the C-terminal end.

A "host cell" is selected from the group consisting of:  
5 fungi, yeasts, and prokaryotes. The micro-organism is more preferably a prokaryote and most preferably a bacterium of the genus *Bifidobacterium* or the species *E. coli*.

10 By "oligosaccharides" is meant an oligosaccharide consisting of at least three sugar molecules. An example of an oligosaccharide, which is not meant to be limiting, is galacto-oligosaccharide. The linkages between the sugar residues of the oligosaccharide comprise but are  
15 not limited to 1-4 and 1-6 bindings.

Incubation of  $\beta$ -galactosidase with lactose takes place in the presence of 0.5-60% lactose, preferably 2-30% lactose and most preferably 2-15% lactose.

20 Conditions of incubating  $\beta$ -galactosidase with lactose are defined by performing the incubation at a temperature between 5 and 75 °C, preferably 15-45 °C, and most preferably at 37 °C. The time required for the incubation  
25 is 1-50 hours, preferably 5-40 hours and most preferably 15-25 hours.

A "comestible product" comprises a product intended for ingestion such as foods, drinks, tablets, and powders.

30

## Examples

### Example 1:

- 5 Isolation and characterisation of transgalactosylating  $\beta$ -galactosidase from *B. bifidum*. *Pst*I digested chromosomal DNA from *B. bifidum* DSM 20215 was ligated into pKS plasmid (Stratagene) using standard procedures. The ligation mixture was transformed into *E. coli* strain
- 10 MT102 defective in LacZ and  $\beta$ -galactosidase.  $\beta$ -galactosidase producing clones were identified as blue colonies on plates containing the chromogenic  $\beta$ -galactosidase substrate X-gal.
- 15 One of the blue colonies contained a plasmid with an insert of approximately 20 kb, pOLGA5. The insert was further subcloned and partly sequenced and an open reading frame encoding a putative  $\beta$ -galactosidase (OLGA5  $\beta$ -galactosidase) was identified (Figure 1). BLAST search
- 20 showed that OLGA5  $\beta$ -galactosidase showed the highest degree of homology with *Streptomyces coelicolor*  $\beta$ -galactosidase (AL133171) and *Thermoanaerobacter ethanolicus* (Y08557) with 38% and 30% identity, respectively. Figure 3 shows an "identity tree" of OLGA5
- 25 and related amino acid sequences.

A detailed analysis of the amino acid sequence of OLGA5  $\beta$ -galactosidase revealed that the enzyme contains a putative signal sequence at its N-terminal and that the

30 open reading frame encodes a polypeptide of 185 kDa which is approximately twice as large as any of the presently known  $\beta$ -galactosidases. Recombinant OLGA5 enzyme produced in *E. coli* was purified and N-terminal amino acid sequencing confirmed, that the signal sequence was

cleaved during expression in *E. coli*. SDS-PAGE confirmed the molecular weight of the OLGA5 polypeptide.

Cellular extracts of recombinant *E. coli* MT102 containing  
5 pOLGA5 were prepared and analysed for  
transgalactosylating activity. Figure 4 shows that OLGA5,  
in addition to lactose hydrolysing activity, also  
exhibited transgalactosylating activity.

## 10 Example 2

Construction of a truncated OLGA5  $\beta$ -galactosidase with  
high transgalactosylase activity. The region of OLGA5  
homologous to other  $\beta$ -galactosidases is located in the N-  
15 terminal end of the protein. The C-terminal half showed  
no homology to any known  $\beta$ -galactosidase. However, a  
sialidase-like galactose-binding domain was observed in  
the C-terminal part. The role of this C-terminal part of  
the OLGA5  $\beta$ -galactosidase was investigated by  
20 construction of truncated deletion mutants. The  
hydrolytic and transgalactosylating activities of the  
resulting recombinant  $\beta$ -galactosidases were analysed.  
Figure 5 shows that it was possible to delete almost one  
third of the OLGA5 enzyme and still retain hydrolytic  
25 activity.

When the transgalactosylating activity was analysed,  
similar results were obtained with extracts from *E. coli*  
containing the plasmids pOLGA5, pOLGA342, and pOLGA345.  
30 However, extracts of cells harbouring pOLGA347 showed an  
increased level of oligosaccharides produced and almost  
no galactose. As shown in Figure 5, an extract containing  
the truncated OLGA347  $\beta$ -galactosidase did hydrolyse  
lactose, but instead of transferring galactose onto



hydroxyl groups in water, the enzyme transferred virtually all galactose molecules onto galactose or glucose (or glycerol; the spot migrating slightly slower than glucose on TLC was shown by NMR to be galactoglycerol - data not shown). In conclusion OLGA347 is a true "transgalactosylase".

### Example 3

10 Characterisation of the transgalactosylating activity of OLGA347. Two methods were used to quantitate the transgalactosylating activity of OLGA347  $\beta$ -galactosidase: TLC analysis of reaction mixtures containing radioactively labelled lactose and HPLC analysis after  
15 enzymatic conversion of unlabeled lactose.

Experiments with radioactivity were carried out with lactose containing the  $^{14}\text{C}$ -label at the C-1 position of glucose. Since the label was in the glucose part of the  
20 disaccharide, only reaction products containing glucose were detected. Figure 7 shows the result of a transgalactosylation experiment with 15% lactose and varying amounts of OLGA347 enzyme. After separation of the reaction mixture by TLC, the plate was scanned and  
25 the radioactive spots were quantitated in a phospho-imager. At low enzyme concentrations (between 0 and 0.2  $\mu\text{l}$  of the extract), the glucose and oligosaccharide levels were almost identical, indicating that all glucose molecules were exploited as substrate in  
30 transgalactosylation reactions. "Free" hydrolysed glucose appeared only at high enzyme concentrations.

In experiments with unlabelled lactose different substrate and enzyme concentrations were examined. Figure 8 shows an experiment in which 10%, 20%, and 40% lactose was used as substrate in enzyme reactions with varying concentrations of OLGA347 enzyme. The reaction mixtures were analysed with HPLC and the concentrations of lactose, glucose, galactose, and galacto-oligosaccharides were calculated. Figure 8 shows that as the enzyme concentrations goes up, the lactose concentration is decreased and the glucose concentration is increased but virtually no "free" galactose is produced, indicating that almost all galactose molecules in lactose are transferred onto another sugar. Calculations of carbohydrate concentrations measured in reactions with low enzyme concentrations, indicated that the ratio between glucose and galactose is approximately 0.1, implying that for every lactose molecule hydrolysed to free galactose and glucose, nine lactose molecules are used in transgalactosylation. As seen in Figure 8, the transgalactosylation reaction is independent of lactose concentration in range from 10% to 40% lactose. The maximal yield of galacto-oligosaccharides produced in transgalactosylation reactions with 10%, 20% or 40% lactose as substrate were 39%, 44% and 37% respectively (mg of oligosaccharides produced per mg lactose added).

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## Claims

1. A DNA sequence which
- 5
- a) encodes a protein with an amino acid sequence as given in SEQ ID NO:2, or
  - b) hybridises under stringent conditions to the sequence of a), or
  - 10 c) is degenerative of the sequence of a) or b).
2. A DNA sequence according to claim 1, wherein the sequence is as given in SEQ ID NO:1 or a fragment thereof.
- 15
3. A DNA sequence according to claim 2, wherein the sequence comprises a sequence from SEQ ID NO:1 which starts with ATG in position 212-214 and ends with TGA in position 5468-5470, or any fragment thereof.
- 20
4. A DNA sequence according to claim 3, wherein the sequence comprises a sequence from SEQ ID NO:1 which starts with ATG in position 212-214 and ends with ATCT in position 3731-3734, or any fragment thereof.
- 25
5. A DNA sequence according to claim 3, wherein the sequence comprises a sequence from SEQ ID NO:1 which starts with GTC in position 308-310 and ends with TGA in position 5468-5470, or any fragment thereof.
- 30

- 5           6. A DNA sequence according to claim 3, wherein the sequence comprises a sequence from SEQ ID NO:1 which starts with GTC in position 308-310 and ends with ATCT in position 3731-3734, or any fragment thereof.
- 10           7. A DNA sequence according to any one of claims 1-6, wherein said sequence comprises nucleotide substitutions, additions or deletions which result in less than 60%, preferably less than 45%, more preferably less than 25% change in the amino acid sequence according to SEQ ID NO:2, or a fragment thereof.
- 15           8. A DNA sequence according to any one of claims 1-5, wherein said sequence comprises nucleotide substitutions, which results in conservative amino acid substitutions.
- 20           9. An enzyme encoded by a DNA sequence of any one of claims 1-8.
10. An enzyme comprising an amino acid sequence according to SEQ ID NO:2, or a fragment thereof.
- 25           11. A  $\beta$ -galactosidase having the sequence as defined in SEQ ID NO:2.
- 30           12. An enzyme according to claim 10 having the sequence as defined in SEQ ID NO:2 from Met (1) to Gly (1752), or a fragment thereof.
13. A mature  $\beta$ -galactosidase according to claim 12.

14. An enzyme according to claim 10 having the sequence as defined in SEQ ID NO:2 from Met (1) to Ile (1174), or a fragment thereof.
- 5 15. A transgalactosylating enzyme according to claim 14.
- 10 16. An enzyme according to claim 14 having the sequence as defined in SEQ ID NO:2 from Ala (33) to Ile (1174), or a fragment thereof.
17. A mature transgalactosylating enzyme according to claim 16.
- 15 18. A transgalactosylating enzyme of any one of claims 14-17 having one or more of the following characteristics:
- 20 a) The ratio of transgalactosylating activity to  $\beta$ -galactosidase activity in a solution of 100 g/L lactose at 37 °C is at least 1:1,
- b) catalyses production of at least 25% galacto-oligosaccharides in batch reaction with a solution of 100 g/L lactose at 37 °C,
- 25 c) catalyses production of galacto-oligosaccharides in batch reaction with a solution of 100 g/L lactose at 37 °C with less than 15% of galactose from the lactose being present in the free form at the reaction time with
- 30 maximum concentration of galacto-oligosaccharide.
19. A recombinant vector comprising a DNA sequence of any one of claims 1-8.



20. A vector of claim 19, wherein said vector is an expression vector.
- 5 21. A host cell comprising a DNA sequence of any one of claims 1-8.
22. A host cell comprising a vector of any one of claims 19-20.
- 10 23. A cell of claims 21-22, wherein said cell is a bacterial cell, a yeast cell, or a fungal cell.
- 15 24. A cell of claim 23, wherein the cell is selected from the group consisting of *Bifidobacterium*, *Lactococcus*, *Lactobacillus*, *Streptococcus*, *Leuconostoc*, *Escherichia*, *Bacillus*, *Streptomyces*, *Saccharomyces*, *Kluyveromyces*, *Candida*, *Torula*, *Torulopsis* and *Aspergillus*.
- 20 25. A cell of claim 24, wherein the cell is selected from the group consisting of *Bifidobacterium breve*, *Bifidobacterium longum*, *Bifidobacterium infantis*, *Bifidobacterium bifidum* and *Lactococcus lactis*.
- 25 26. Use of a cell of any one of claims 21-25 for producing a product selected from the group consisting of yoghurt, cheese, fermented milk product, dietary supplement and probiotic
- 30 comestible product.
27. A dairy product comprising a cell of any one of claims 21-25.

28. Use of a transgalactosylating enzyme of any one of claims 14-18 or a cell of any one of claims 21-25, for producing galacto-oligosaccharides.
- 5 29. Use of a transgalactosylating enzyme of any one of claims 14-18 or a cell of any one of claims 21-25, for producing galacto-oligosaccharides to be part of a product selected from the group consisting of yoghurt, cheese, fermented dairy products, dietary  
10 supplements and probiotic comestible products.
30. Use of a transgalactosylating enzyme of any one of claims 14-18 or a cell of any one of claims 21-25, for producing galacto-oligosaccharides to enhance  
15 the growth of *Bifidobacterium*.
31. Use of a transgalactosylating enzyme of any one of claims 14-18 or a cell of any one of claims 21-25, for producing galacto-oligosaccharides to enhance  
20 the growth of *Bifidobacterium* in a mixed culture fermentation.
- 32.A process for producing a transgalactosylating enzyme of any one of claims 14-18, comprising  
25 culturing a cell of any one of claims 21-25 in a suitable culture medium under conditions permitting expression of said enzyme, and recovering the resulting enzyme from the culture.
- 30 33.A process for producing galacto-oligosaccharides, comprising contacting of an enzyme of any one of claims 14-18 or a cell of any one of claims 21-25 with a solution of lactose.

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1 ATGCGTTGCGTTGCGATTTTTCCGGCCCTGTATGGGGGATACAGGATTGGCGATGGCGACACGCCGTTTTTGTAAATGGC  
81 ATTTACATGAAATACAGGTAATGAGATATCATTTCTCATGATCACCGTGTGGATATCGCATTGGTGGTATACACTAACAG  
161 CAACAGAGCGGCGCGGCAGGCGCTCGTGGATTCAATGAAGAAGGAACGTTTATGGCAGTTTCGCAGACTTGGTGGCCGCAT  
M A V R R L G G R I  
241 CGTGGCTTTTCGCCGCCACAGTGGCCTTGTCAATACCGTTAGGGTTGTTAACAATTCAGCGTGGGCGGTTCGAGGACGCCA  
V A F A A T V A L S I P L G L L T N S A W A V E D A  
321 CCCGATCCGACTCCACCACGCAGATGAGCTCCACGCCGAGGTGGTCTACTCCAGCGCCGTGGATTCCAAGCAGAATCGC  
T R S D S T T Q M S S T P E V V Y S S A V D S K Q N R  
401 ACCTCGGATTTTCGACGCCAAGTGAAGTTTCATGCTGTCCGATTCCGTGCAGGCGCAGGATCCGGCGTTTCGACGATTCCGC  
T S D F D A N W K F M L S D S V Q A Q D P A F D D S A  
481 CTGGCAGCAGGTTCGACCTGCCGCATGACTACAGCATCACGCAGAAGTATTCGAGAGCAACGAGGCCGAAAGCGCATACC  
W Q Q V D L P H D Y S I T Q K Y S Q S N E A E S A Y  
561 TTCCCGCGGCGCACCGGCTGGTACCGCAAGTCCTTCACCATCGACCGGGACCTCGCCGCGCAAGCGCATCGCCATCAACTTC  
L P G G T G W Y R K S F T I D R D L A G K R I A I N F  
641 GACGCGGTGTACATGAACGCCACCGTCTGGTTCACCGCGTCAAGCTCGGCACCCATCCGTACGGCTACTCGCCGTTCTC  
D G V Y M N A T V W F N G V K L G T H P Y G Y S P F S  
721 CTTGACCTGACCGGCAACGCCAAGTTCGGTGGGAGAACACCATCGTCTCAAGGTTCGAGAACAGGCTGCCGTCCAGCC  
F D L T G N A K F G G E N T I V V K V E N R L P S S  
801 GCTGGTACTCCGGCTCCGGCATCTACCGCGACGTCACCCCTCACCGTCACCGACGGCGTGCACGTCGGCAATAACGGCGTG  
R W Y S G S G I Y R D V T L T V T D G V H V G N N G V  
881 GCCATCAAGACCCCGAGCCTCGCCACCCAAAACGGCGGCGACGTGACGATGAACCTCACCAAGGTTCGCCAACGACAC  
A I K T P S L A T Q N G G D V T M N L T T K V A N D T  
961 CGAGGCGCGGCGGAACATCACCTCAAGCAGACCGTGTTCCTCCCAAGGGAGGCAAGACCGACGCGCCCATCGGCACCGTCA  
E A A A N I T L K Q T V F P K G G K T D A A I G T V  
1041 CCACCGCATCCAAGTCCATCGCGGCGGTGCCAGCGCGGACGTGACCTCCACGATCACCGCCGCTTCGCCCAAGCTGTGG  
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1121 AGCATCAAGAACCCGAACCTGTACACCGTGGCGACCGAAGTGCTCAACGGCGGCAAGGTGCTCGACACTTACGACACCGA  
S I K N P N L Y T V R T E V L N G G K V L D T Y D T E  
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S M H H D Q G S L G A V A N R R A I E R Q V E I L Q K  
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M G V N S I R T T H N P A A K A L I D V C N E K G V L  
1441 CGTGGTTCGAAGAGGTCTTCGACATGTGGAACCGGTTCGAAGAACGGCAACACCGAGGATTACGGCAAGTGGTTCGGCCAGG  
V V E E V F D M W N R S K N G N T E D Y G K W F G Q  
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A I A G D N A V L G G D K D E T W A K F D L T S T I N  
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F P A T S A K L V A W T K A A D S T R P M T Y G D N  
1761 AGATCAAGGCCAAGTGAACGAGTGAACACCATGGGCGACAACCTGACCGCCAACGGCGGCGTGGTTCGGCACCAACTAC  
K I K A N W N E S N T M G D N L T A N G G V V G T N Y  
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S D G A N Y D K I R T T H P S W A I Y G S E T A S A I  
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N S R G I Y N R T T G G A Q S S D K Q L T S Y D N S  
2001 CAGTCCGCTGGGGCGCCGTCGCCAGCTCCGCTGGTACGACGTGGTTCAGCGCGATTTCGTCGCCGGCACATACGTGTGG  
A V G W G A V A S S A W Y D V V Q R D F V A G T Y V W  
2081 ACCGGCTTCGACTACCTCGGCGAACCCACCCCGTGAACGGCACCGGCTCCGGCGCCGTGGGCTCCTTGGCCGTTCGCCGA  
T G F D Y L G E P T P W N G T G S G A V G S L A V A E  
2161 AGAACTCGTACTTCGGCATCGTCGACACCGCAGGCTTCCCGAAGACACCTATTACTTCTATCAGAGCCAGTGAACGACG  
E L V L R H R R H R R L P E D T Y Y F Y Q S Q W N D  
2241 ACGTGCACACGCTGCACATCCTCCCGCATGGAACGAGAACGTCGTCGCCAAGGGCTCCGGCAACAACGTGCCGGTTCGTC  
D V H T L H I L P A W N E N V V A K G S G N N V P V V

Fig. 1

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2321 GTCTACACCGACGCGGCCAAGGTCAAGCTGTACTTCACACCGAAGGGCAGTACCGAAAAGCGACTGATCGGAGAGAAGTC  
V Y T D A A K V K L Y F T P K G S T E K R L I G E K S

2401 CTTACCAAGAAGACCACCGCGGGCCGGATACACCTATCAGGTCTACGAGGGCTCCGACAAGGACTCCACCGCCCACAAGA  
F T K K T T A A G Y T Y Q V Y E G S D K D S T A H K

2481 ACATGTACCTGACCTGGAACGTGCCGTGGGCGGAGGGCACCATCTCCGCCGAAGCATACGACGAGAACACAGGCTGATC  
N M Y L T W N V P W A E G T I S A E A Y D E N N R L I

2561 CCGAGGGGTCCACCGAGGGCAACGCGTGGGTGACCACCACCGGCAAGGCCGCGAAGCTTAAAGCCGATGCCGACCGCAA  
P E G S T E G N A S V T T T G K A A K L K A D A D R K

2641 GACGATCACCGCGGACGGCAAGGACCTGTCTGATCATCGAGGTGACGTGACCGACGCCAACGGCCATATCGTCCCGGATG  
T I T A D G K D L S Y I E V D V T D A N G H I V P D

2721 CCGCCAACCGCTCACCTTCGACGTCAAGGGCGCCGGCAAACCTGGTGGCGTGCACAACGGCAGCTCGCCGGATCACGAC  
A A N R V T F D V K G A G K L V G V D N G S S P D H D

2801 TCCTATCAGGCGGACAACCGCAAGGCGTTCAGCGGCAAGGTGCTCGCCATCGTCCAGTCCACCAAGGAGGGCGGCGAGAT  
S Y Q A D N R K A F S G K V L A I V Q S T K E A G E I

2881 CACCGTCACCGCCAAGGCCGACGGTCTGCAATCATCCACAGTGAAGATCGCCACCACCGCCGTCCCGGACCGACCGG  
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2961 AGAAGACGGTCCGCGAGCTTCTACTACTCGCGCACTACTACGTCAAGACCGGCAACAAGCCGATTCTGCCGAGTGATGTC  
E K T V R S F Y Y S R N Y Y V K T G N K P I L P S D V

3041 GAGGTGCGCTACTCCGACGGCAGCTCGGACCGTCAAGACGTACATGGGACGCGAGTCAGCGACGACCGAGATCGCCAAGGC  
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3121 CGGTTCGTTTCAGCGTGGCGGGCACGGTCCGCGGGCAGAAGATCTCCGTGCGCGTGACGATGATCGACGAGATCGGTGCGC  
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3201 TGCTCAACTATTTCGGCCAGCACACCGGTCGGCACGCCCCGCGTGTGCTGGCTCGCGTCCGGCCGTGCTGCCCGACGGC  
L L N Y S A S T P V G T P A V L P G S R P A V L P D G

3281 ACCGTGACCGCGCGAAGTTCGCGTCCACTGGACCAAGCCCCGCGACACCGTGTACACACGGCCGGCACCGTCAAGGT  
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3361 CCGCGCACCGCCACCGTCTTCGGCAAGGAGTTCAAGGTACCGCGACGATTTCGCGTGCAGCGGTGCGAGGTACCATCG  
P G T A T V F G K E F K V T A T I R V Q R S Q V T I

3441 GCAGCAGCGTCTCCGGCAATGCGCTGCGCCTGACTCAGAACATCCCCGCGGACAAGCAGTCCGACACGCTGGACGCCATC  
G S S V S G N A L R L T Q N I P A D K Q S D T L D A I

3521 AAGGACGGCTCCACGACCGTCGACGCCAATACCGGGCGGCGCGGAACCCGTCAGCATGGACCAACTGGGCGTACTCGAA  
K D G S T T V D A N T G G G A N P S A W T N W A Y S K

3601 GGCGGGCCACAACACCGCCGAGATCACCTTCGAGTACGCGACCGAGCAGCAGCTCGGCCAGATTGTCATGTACTTCTTCC  
A G H N T A E I T F E Y A T E Q Q L G Q I V M Y F F

3681 GCGACAGCAACGCGGTGAGGTTCCCCGACGCGCGGCAAGACGAAGATCCAGATCTCCGCGGACGGCAAGAAGTGGACGGAT  
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3761 CTCGCTGCCACGGAGACCATCGCGGGCCAGGAGTCGTCCGACCGAGTCAAGCCGTACACCTATGACTTCGCTCCGGTGGG  
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3921 TCGAGCTGAAGACCGCGACCGAGCAAGTTCGTCACGAACACGTCCGCGCGCTCTCGTTCGCTGACAGTGAACGGCAGCAAG  
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4001 GTCTCCGACTCCGTGCTCGCCGCGGGCTCCTACAACACGCGCGGATCATCGCGGACGTCAAAGCCGAGGGCGAAGGCAA  
V S D S V L A A G S Y N T P A I I A D V K A E G E G N

4081 CGCCAGCGTCACCGTGTGCGCGCGCACGACAACGTGATCCGCGTGATCACCGAGTCCGAGGACCACGTACCGCGCAAGA  
A S V T V L P A H D N V I R V I T E S E D H V T R K

4161 CCTTCACCATCAACCTGGGCACGGAGCAGGAATTCCCCGAGACTCCGATGAACGCGACTACCGGGCCCGGACATGACG  
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4241 GTCACCGTGGGCAGCGAACAGACGTCCGGCACCGCGACCGAAGGCCCGAAGAAATTCGCGGTGACGGCAACACCAGCAC  
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4321 GTACTGGCATTCCAACCTGGACGCGCCACCACCGTGAACGACCTGTGGATCGCCTTCGAGCTCCAGAAACCCACCAAGCTCG  
Y W H S N W T P T T V N D L W I A F E L Q K P T K L

4401 ACGCGCTGCGCTACCTGCCGCGCCCCGCGGGCAGCAAGAAGGCTCCGTACCGAATACAAGGTTTCAGGTACCGATGAC  
D A L R Y L P R P A G S K N G S V T E Y K V Q V S D D

4481 GGCACCAACTGGACCGACGCGGGCTCCGGCACATGGACCACCGATTACGGCTGGAAGCTCGCCGAGTTCAATCAGCCGGT  
G T N W T D A G S G T W T T D Y G W K L A E F N Q P V

Fig. 1 (continued)

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4561 GACCACCAAGCACGTGCGGGCTCAAGGCCGTCCACACCTATGCGGATTCCGGCAACGACAAGTTCATGTCCGCCTCCGAAA  
T T K H V R L K A V H T Y A D S G N D K F M S A S E

4641 TCCGCCTGCGCAAGGCCGTGACACCACCGACATCAGCGGCGCGACCGTGACCGTGCCCGCCAAGCTGACCGTCGACCGG  
I R L R K A V D T T D I S G A T V T V P A K L T V D R

4721 GTGGACGCCGACCATCCCGCCACCTTCGCCACGAAGGACGTGACGGTGACGTTGGGCGACGCCACGCTGCGCTACGGCGT  
V D A D H P A T F A T K D V T V T L G D A T L R Y G V

4801 GGACTACCTGCTCGACTACGCGGGCAACACCGCCGTCCGCAAGGCCACGGTGACCGTGCGCGGCATCGACAAGTACTCCG  
D Y L L D Y A G N T A V G K A T V T V R G I D K Y S

4881 GCACCGTCGCCAAGACGTTACCATCGAACTGAAGAACGCCCCGGCGCCGGAACCGACGCTGACCTCGGTGAGCGTCAAG  
G T V A K T F T I E L K N A P A P E P T L T S V S V K

4961 ACCAAGCCTTCCAAGCTGACCTATGTGGTTCGGCGACGCGTTCGACCCGGCAGGACTGGTGCTGCAGCACGACAGACAGGC  
T K P S K L T Y V V G D A F D P A G L V L Q H D R Q A

5041 CGATCGCCCCCACAGCCACTTGTGGAGAACAGGCCGACGAACGCGGACTGACGTGCGGAACGCGATGCGATCGCGTTG  
D R P P Q P L V G E Q A D E R G L T C G T R C D R V

5121 AACAGCTGCGCAAACACGAGAATCGTGAAGCCCATCGTACGGGCCTCGATCATCTGGAATTCGTGGGTGCCGCCGATGGA  
E Q L R K H E N R E A H R T G L D H L E F V G A A D G

5201 GCGGTTCGGTGAACAGGCCACCTTCAAGGTGCATGTCCATGCCGATCAAGGTGACGGCCGCCATGATGATGCCGATGAACG  
A V G E Q A T F K V H V H A D Q G D G R H D D A D E R

5281 CGATATCGATCCACATGTCCCTGTGATCAGCGGGTCCGTGAGCTTGC CGGGCTGCGTGCCATCACGTCATCGGTCTGC  
D I D P H V P V D H A V G E L A R A A C H H V I G L

5361 GGGTCGACACCCATCGCCTCAAGGCATCCGGCTTCCAGATCCCCGCCGACGACATGGCCGAGATCGACCGCATCACCGGC  
R V D T H R L K A S G F Q I P A D D M A E I D R I T G

5441 TTCCACCGCTTCGAGCGCCACGTCCGCTGACGTGATTGGGCTTCCCCGCTGTCTGGTGCCGGCTCGCGA  
F H R F E R H V G Z

Fig. 1 (continued)

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L35444	RFLAASQAY--LDALAKQVQPLLN-HNGGP-II-AVQVE-NEYGSYAD
M13466	HYCPNHPQL--ITHIKRLVRAIAERYKNHPALK-MWHVN-NEYACHVS
U17417	TISSSAWYYSVGQYAAKMTALAERYKDHPALA-LWHVD-NELGCHVS
B05040	HWRATSPVF--LDYALNLCRKMAEHYKDNPYVV-SWHVS-NEYGCHNR
OLGA88	HWRPTSPVF--REYALRLCRAMAEHYRDNPYVV-AWHVS-NEYGCHNR
L03424	NSCPNSPTY--RKYSEKIADKLAERYKDHPAVL-VWHIS-NEYGDCY
L03425	NHCYTSPVY--REKVTAINTKLAERYSDHPAVI-GWHIS-NEFGGDCH
D49537	NHCYTSPYI--REKIAIDRLLAERYKDHPALI-LWHIS-NEFEGQCY
L20757	RWGGME-TG--GNPERPPHRSSATG--TTRLISY-IWGVIRINESQDSHD
M57579	QYIGNS-EW--KKVAEQNLREMITRDWNHPSII-LWGVIRINESQDDDA
Y08557	QHIGDE-NW--KNIKENLKEMLRDRNHPCIF-MWGVIRINERLDDHD
OLGA5	AVLGCDKDE--TWAKFD-LTSTINRDRNAPSVI-MWSLG-NEMMEGSI
M63636	NIPASEPEW--LPACLDRAANNMFQDRKNHASVI-IWSCG-NESYAGKD
M35107	NVPGSLPQW--QAAVLDRASSMVERDKNHPSVI-IWSCG-NESYAGED
M92281	NVPGDNPHW--PAAVIDRARSNEYWFKNHPSII-FWSLG-NESYAGED
X82287	NVPGSYDEW--EAATLDRARTNFBTFKNHVSII-FWSLG-NESYAGSV
M23530	NVPGDDQHW--LGASLSRVKNMMARDKNHASII-IWSLG-NESYAGTV
AJ242596	IVPGSKREW--EGACVDRVNSMMRRDYNHPSVI-IWSLG-NESYVGDV
OLGA2	SVPGDDEAW--LGACIDRLDSMILRDRNHPSVI-VWSLG-NESYAGEV
U62625	CYFARDPLF--KKAILDRQQANVERDKNRTSII-IWSLG-NEAGYGAN
Y14599	NIIADDSKF--ETAIIERIEASIMPLKNYSSIV-SWSLG-NESGFGKN
U08186	VTLANRWEW--EKAHFDRIKRMVERDKNHPSII-FWSLG-NEAGDGVN
OLGA1	RPIADNPAW--IAPTVDRAQRSVERDKNHASII-FWSMG-NECAYGCT
M11441	NRLSDDPAW--LPAFSARVTRMVQSNRNHPSII-IWSLG-NESGGGCGN
U60828	NRLTNDPTY--LPLMSERVTRMVMRDRNHPSII-IWSLG-NESGYGSN
J01636	NRLTDDPRW--LPAMSERVTRMVQRDRNHPSVI-IWSLG-NESGHGAN
D42077	SRLADDPRW--LPAMSERVTRMVQRDRNHPSII-IWSLG-NESGHGAN
D37882 (P)	EGLHEDGDFLTHEKMDDFVEYADYCFKEFPEVK-YWITI-NEIRSVAV
J03479 (P)	EVLHKDGDFLNRKTIDYFVDYAEYCFKEFPEVK-YWTTT-NEIGPIGD
L18993 (P)	EALHSNGDFLNRENIEHFVNIAEFCFKEFSEVN-YWTTT-NEIGPIGD
M28357 (P)	EALHSNGDFLNRENIEHFIDYAAFCFEEFPEVN-YWTTT-NEIGPIGD
M34696	GDFTGPGSWLSTRTVYEFARFSAYIAWKFDLVDDEYSTM-NEPNVVGG
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Fig. 2



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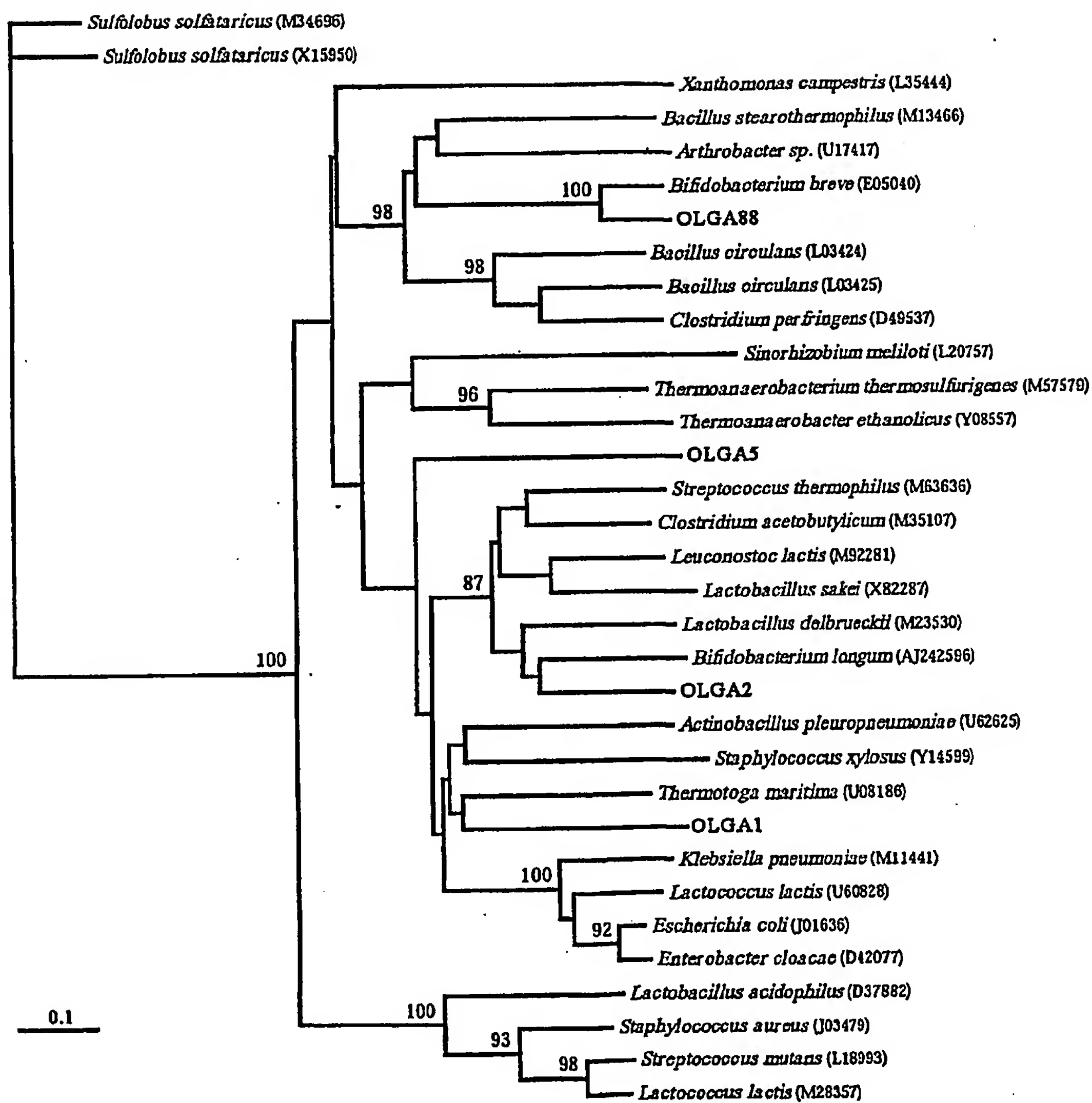


Fig. 3

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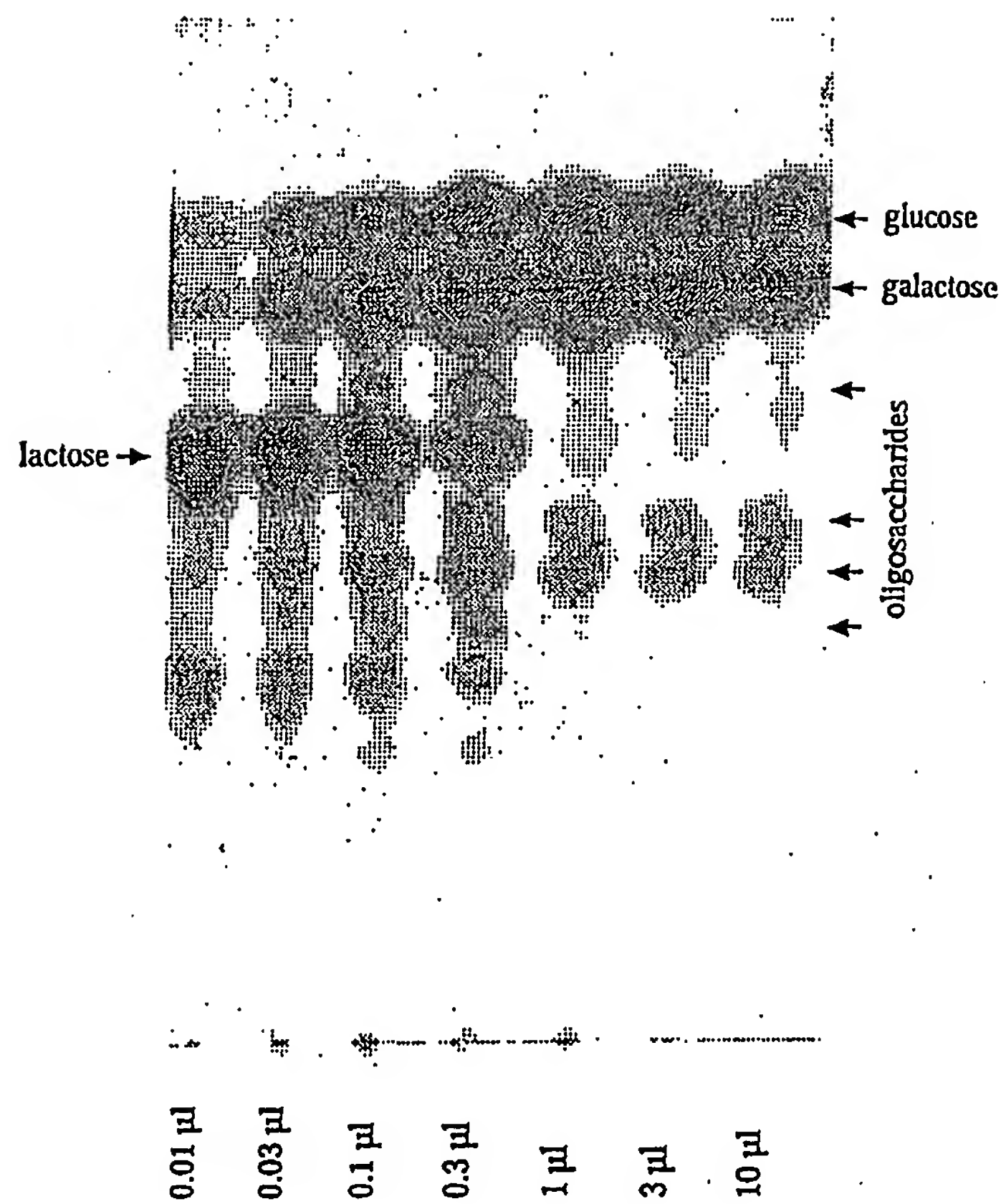


Fig. 4

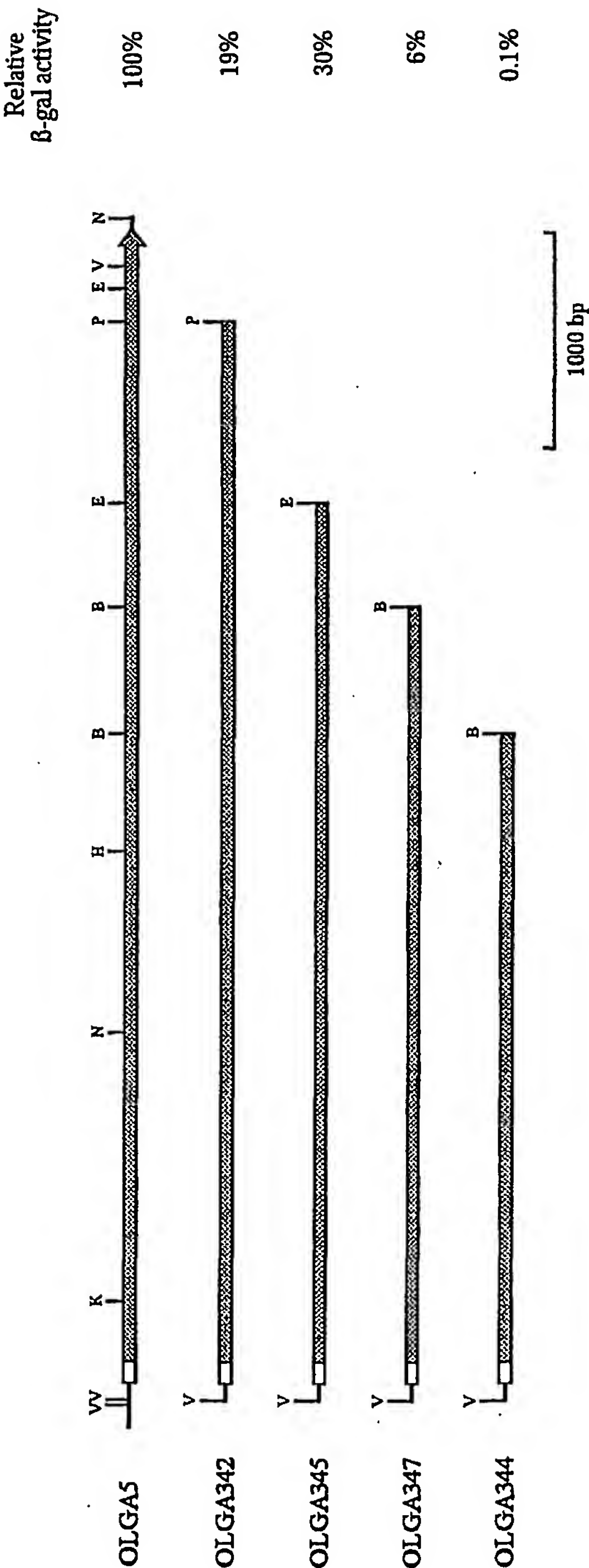


Fig. 5

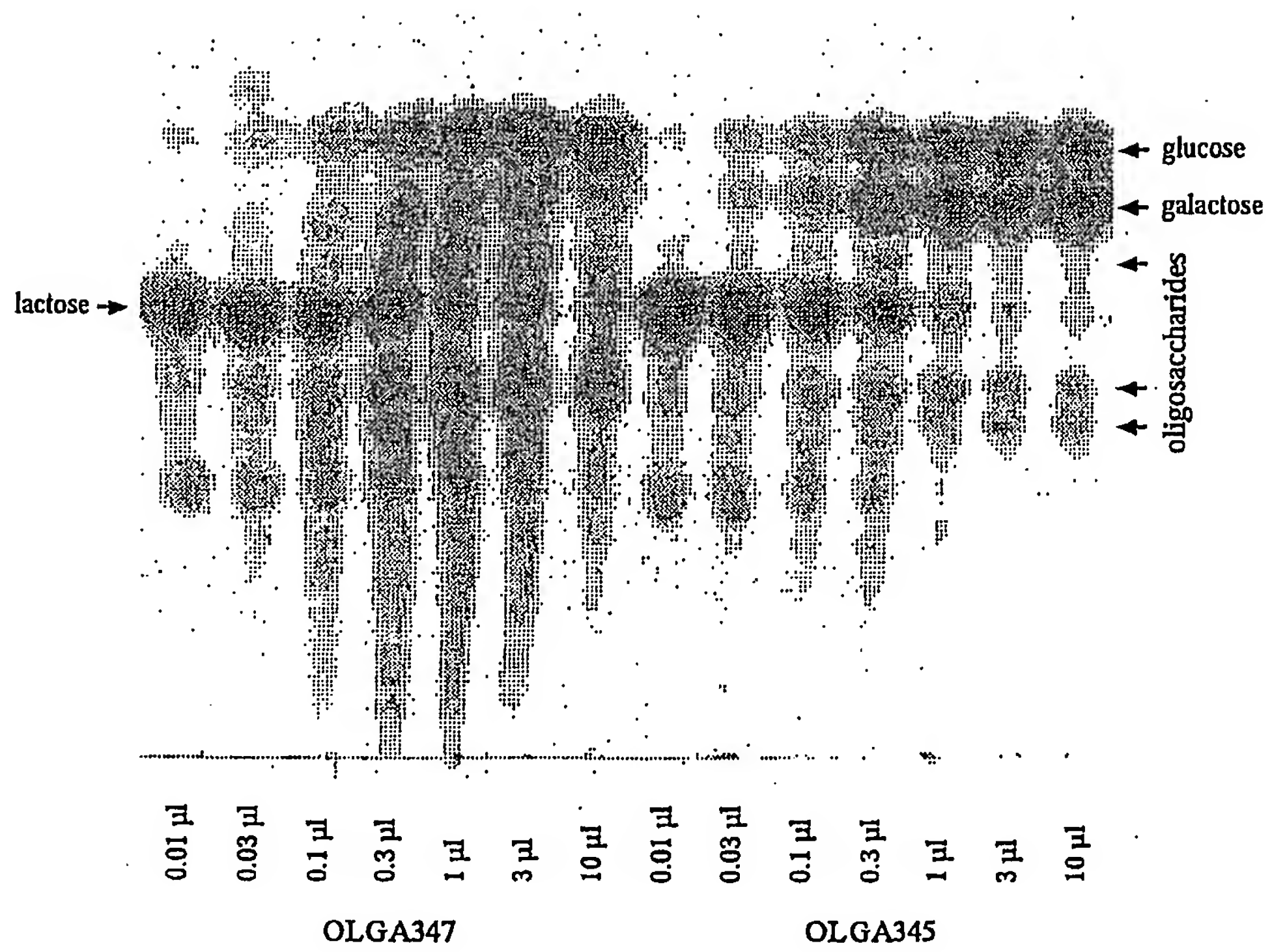
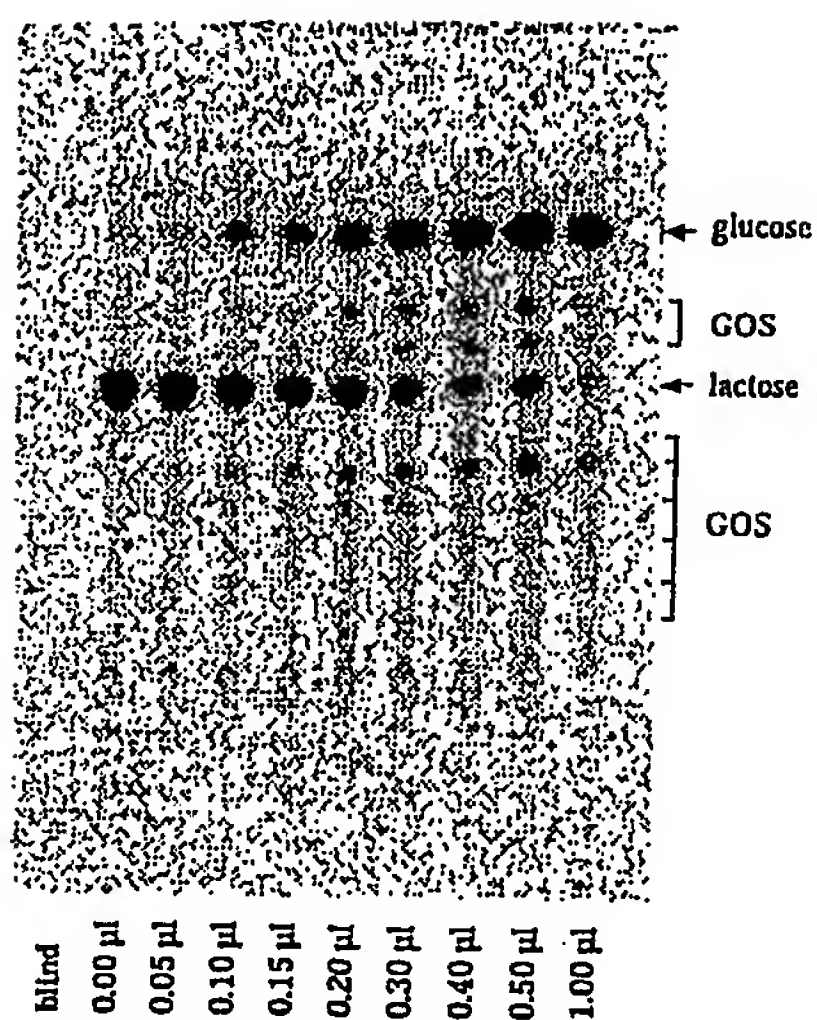


Fig. 6

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(A)



(B)

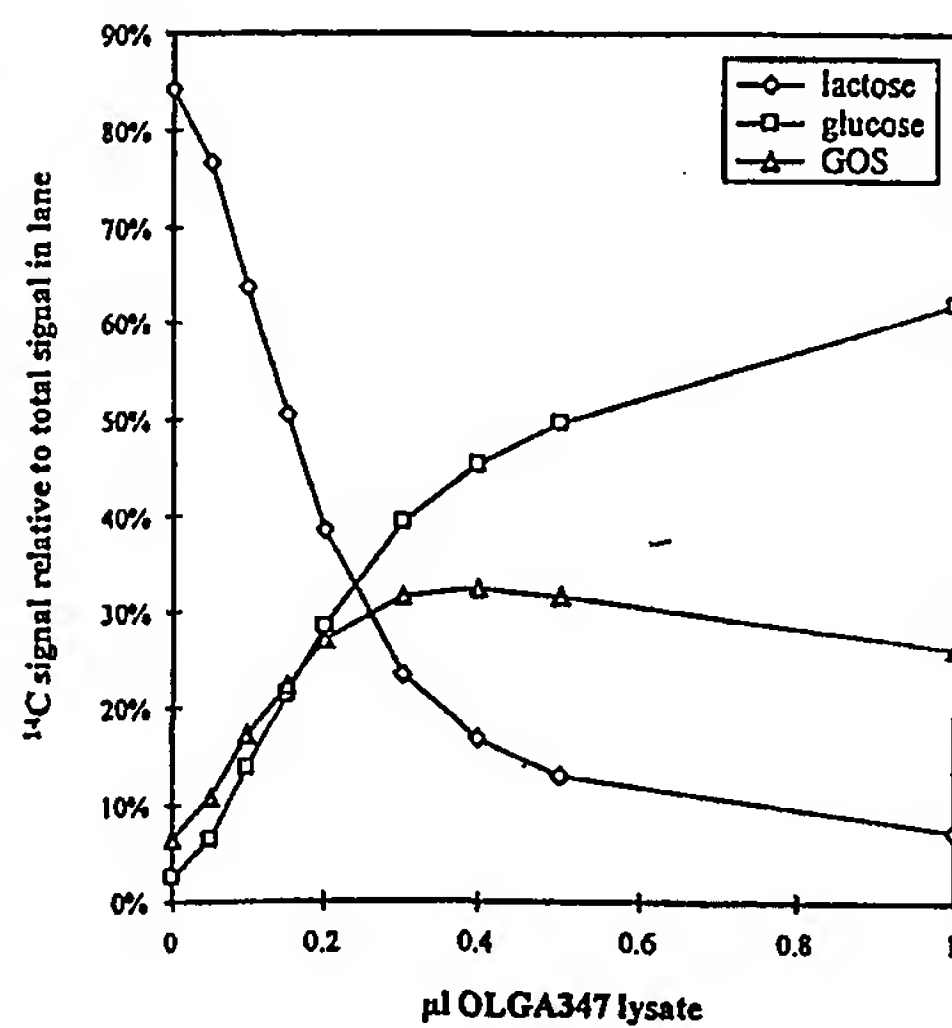


FIG. 7

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**(A)** Reaction with 10% lactose.

	0 $\mu$ l	0.1 $\mu$ l	0.2 $\mu$ l	0.4 $\mu$ l	0.8 $\mu$ l	1.5 $\mu$ l	3 $\mu$ l	6 $\mu$ l
lactose	112.38	105.87	101.35	92.52	75.56	51.82	34.04	30.08
glucose	0	1.52	2.85	6.11	11.53	20.66	30.16	36.92
galactose	0	0.19	0.30	0.66	1.30	2.16	3.80	5.58

**(B)** Reaction with 20% lactose.

	0 $\mu$ l	0.1 $\mu$ l	0.2 $\mu$ l	0.4 $\mu$ l	0.8 $\mu$ l	1.5 $\mu$ l	3 $\mu$ l	6 $\mu$ l
lactose	235.65	217.58	205.30	177.70	137.27	93.78	66.24	61.69
glucose	0	2.95	6.48	13.93	29.57	45.99	61.06	73.06
galactose	0	0.34	0.48	0.78	1.96	3.07	4.87	6.95

**(C)** Reaction with 40% lactose.

	0 $\mu$ l	0.1 $\mu$ l	0.2 $\mu$ l	0.4 $\mu$ l	0.8 $\mu$ l	1.5 $\mu$ l	3 $\mu$ l	6 $\mu$ l
lactose	426.47	395.16	370.29	308.07	224.08	174.88	136.73	121.29
glucose	0	7.96	17.51	37.96	63.42	93.99	123.99	144.27
galactose	0	0.65	0.97	1.48	2.94	4.11	6.84	8.89

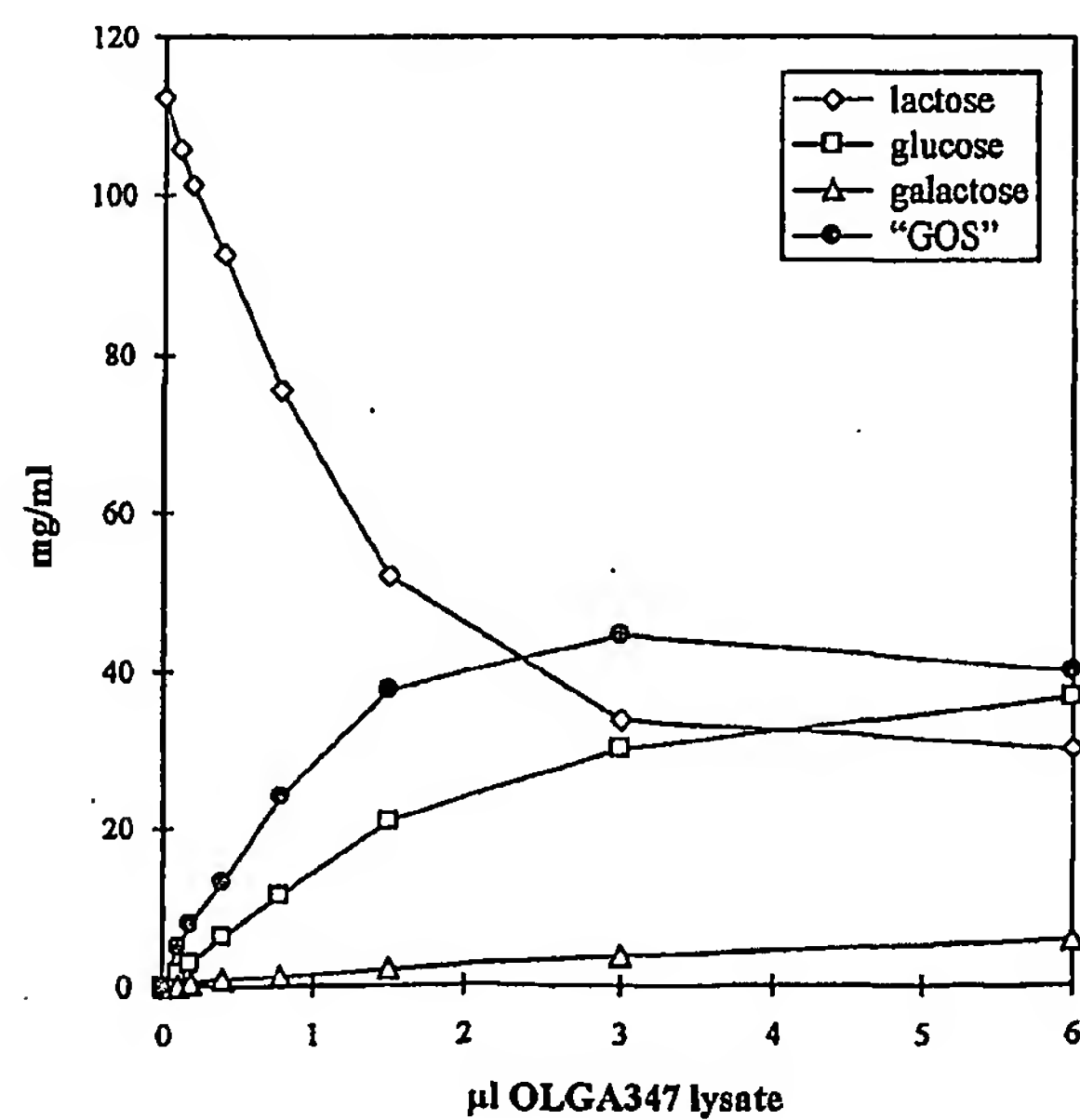
**(D)** Plot of reaction with 10% lactose.

Fig. 8



## SEQUENCE LISTING

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 tcaccgtgtg gatatcgcat tgggtgcgtat acactaacag caacagagcg gcgcggcagg 180  
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 Gly Leu Leu Thr Asn Ser Ala Trp Ala Val Glu Asp Ala Thr Arg Ser  
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 Asp Ser Thr Thr Gln Met Ser Ser Thr Pro Glu Val Val Tyr Ser Ser  
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 Lys Phe Met Leu Ser Asp Ser Val Gln Ala Gln Asp Pro Ala Phe Asp  
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	Asp Thr Glu Tyr Gly Phe Arg Trp Thr Gly Phe Asp Ala Thr Ser Gly	
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	Thr Leu His Ile Leu Pro Ala Trp Asn Glu Asn Val Val Ala Lys Gly	
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	Ser Gly Asn Asn Val Pro Val Val Val Tyr Thr Asp Ala Ala Lys Val	
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10	Ile	Gly	Thr	Val	Thr	Thr	Ala	Ser	Lys	Ser	Ile	Ala	Ala	Gly	Ala	Ser
			275				280						285			
	Ala	Asp	Val	Thr	Ser	Thr	Ile	Thr	Ala	Ala	Ser	Pro	Lys	Leu	Trp	Ser
		290					295					300				
	Ile	Lys	Asn	Pro	Asn	Leu	Tyr	Thr	Val	Arg	Thr	Glu	Val	Leu	Asn	Gly
	305					310					315					320
15	Gly	Lys	Val	Leu	Asp	Thr	Tyr	Asp	Thr	Glu	Tyr	Gly	Phe	Arg	Trp	Thr
					325					330					335	
	Gly	Phe	Asp	Ala	Thr	Ser	Gly	Phe	Ser	Leu	Asn	Gly	Glu	Lys	Val	Lys
			340						345					350		
20	Leu	Lys	Gly	Val	Ser	Met	His	His	Asp	Gln	Gly	Ser	Leu	Gly	Ala	Val
		355					360						365			
	Ala	Asn	Arg	Arg	Ala	Ile	Glu	Arg	Gln	Val	Glu	Ile	Leu	Gln	Lys	Met
		370					375					380				
	Gly	Val	Asn	Ser	Ile	Arg	Thr	Thr	His	Asn	Pro	Ala	Ala	Lys	Ala	Leu
	385					390					395					400
25	Ile	Asp	Val	Cys	Asn	Glu	Lys	Gly	Val	Leu	Val	Val	Glu	Glu	Val	Phe
				405						410					415	
	Asp	Met	Trp	Asn	Arg	Ser	Lys	Asn	Gly	Asn	Thr	Glu	Asp	Tyr	Gly	Lys
			420						425					430		
30	Trp	Phe	Gly	Gln	Ala	Ile	Ala	Gly	Asp	Asn	Ala	Val	Leu	Gly	Gly	Asp
		435						440					445			
	Lys	Asp	Glu	Thr	Trp	Ala	Lys	Phe	Asp	Leu	Thr	Ser	Thr	Ile	Asn	Arg
		450					455					460				
	Asp	Arg	Asn	Ala	Pro	Ser	Val	Ile	Met	Trp	Ser	Leu	Gly	Asn	Glu	Met
	465					470					475					480
35	Met	Glu	Gly	Ile	Ser	Gly	Ser	Val	Ser	Gly	Phe	Pro	Ala	Thr	Ser	Ala
				485						490					495	
	Lys	Leu	Val	Ala	Trp	Thr	Lys	Ala	Ala	Asp	Ser	Thr	Arg	Pro	Met	Thr
			500						505					510		
40	Tyr	Gly	Asp	Asn	Lys	Ile	Lys	Ala	Asn	Trp	Asn	Glu	Ser	Asn	Thr	Met
		515						520					525			
	Gly	Asp	Asn	Leu	Thr	Ala	Asn	Gly	Gly	Val	Val	Gly	Thr	Asn	Tyr	Ser
		530					535					540				
	Asp	Gly	Ala	Asn	Tyr	Asp	Lys	Ile	Arg	Thr	Thr	His	Pro	Ser	Trp	Ala
	545					550					555					560
45	Ile	Tyr	Gly	Ser	Glu	Thr	Ala	Ser	Ala	Ile	Asn	Ser	Arg	Gly	Ile	Tyr
				565						570					575	
	Asn	Arg	Thr	Thr	Gly	Gly	Ala	Gln	Ser	Ser	Asp	Lys	Gln	Leu	Thr	Ser
			580						585					590		
50	Tyr	Asp	Asn	Ser	Ala	Val	Gly	Trp	Gly	Ala	Val	Ala	Ser	Ser	Ala	Trp
		595						600					605			
	Tyr	Asp	Val	Val	Gln	Arg	Asp	Phe	Val	Ala	Gly	Thr	Tyr	Val	Trp	Thr
		610					615					620				
	Gly	Phe	Asp	Tyr	Leu	Gly	Glu	Pro	Thr	Pro	Trp	Asn	Gly	Thr	Gly	Ser
	625					630					635					640
55	Gly	Ala	Val	Gly	Ser	Leu	Ala	Val	Ala	Glu	Glu	Leu	Val	Leu	Arg	His
				645						650					655	
	Arg	Arg	His	Arg	Arg	Leu	Pro	Glu	Asp	Thr	Tyr	Tyr	Phe	Tyr	Gln	Ser
			660						665					670		
60	Gln	Trp	Asn	Asp	Asp	Val	His	Thr	Leu	His	Ile	Leu	Pro	Ala	Trp	Asn
		675						680					685			
	Glu	Asn	Val	Val	Ala	Lys	Gly	Ser	Gly	Asn	Asn	Val	Pro	Val	Val	Val
		690					695					700				
	Tyr	Thr	Asp	Ala	Ala	Lys	Val	Lys	Leu	Tyr	Phe	Thr	Pro	Lys	Gly	Ser
	705					710					715					720

10

	Thr	Glu	Lys	Arg	Leu	Ile	Gly	Glu	Lys	Ser	Phe	Thr	Lys	Lys	Thr	Thr
					725					730					735	
	Ala	Ala	Gly	Tyr	Thr	Tyr	Gln	Val	Tyr	Glu	Gly	Ser	Asp	Lys	Asp	Ser
				740					745					750		
5	Thr	Ala	His	Lys	Asn	Met	Tyr	Leu	Thr	Trp	Asn	Val	Pro	Trp	Ala	Glu
			755				760						765			
	Gly	Thr	Ile	Ser	Ala	Glu	Ala	Tyr	Asp	Glu	Asn	Asn	Arg	Leu	Ile	Pro
		770				775						780				
10	Glu	Gly	Ser	Thr	Glu	Gly	Asn	Ala	Ser	Val	Thr	Thr	Thr	Gly	Lys	Ala
	785					790					795					800
	Ala	Lys	Leu	Lys	Ala	Asp	Ala	Asp	Arg	Lys	Thr	Ile	Thr	Ala	Asp	Gly
					805					810					815	
	Lys	Asp	Leu	Ser	Tyr	Ile	Glu	Val	Asp	Val	Thr	Asp	Ala	Asn	Gly	His
				820					825					830		
15	Ile	Val	Pro	Asp	Ala	Ala	Asn	Arg	Val	Thr	Phe	Asp	Val	Lys	Gly	Ala
				835				840						845		
	Gly	Lys	Leu	Val	Gly	Val	Asp	Asn	Gly	Ser	Ser	Pro	Asp	His	Asp	Ser
		850					855					860				
20	Tyr	Gln	Ala	Asp	Asn	Arg	Lys	Ala	Phe	Ser	Gly	Lys	Val	Leu	Ala	Ile
	865					870					875					880
	Val	Gln	Ser	Thr	Lys	Glu	Ala	Gly	Glu	Ile	Thr	Val	Thr	Ala	Lys	Ala
					885					890					895	
	Asp	Gly	Leu	Gln	Ser	Ser	Thr	Val	Lys	Ile	Ala	Thr	Thr	Ala	Val	Pro
				900					905					910		
25	Gly	Thr	Ser	Thr	Glu	Lys	Thr	Val	Arg	Ser	Phe	Tyr	Tyr	Ser	Arg	Asn
			915					920						925		
	Tyr	Tyr	Val	Lys	Thr	Gly	Asn	Lys	Pro	Ile	Leu	Pro	Ser	Asp	Val	Glu
		930					935					940				
30	Val	Arg	Tyr	Ser	Asp	Gly	Thr	Ser	Asp	Arg	Gln	Asn	Val	Thr	Trp	Asp
	945					950					955					960
	Ala	Val	Ser	Asp	Asp	Gln	Ile	Ala	Lys	Ala	Gly	Ser	Phe	Ser	Val	Ala
					965					970					975	
	Gly	Thr	Val	Ala	Gly	Gln	Lys	Ile	Ser	Val	Arg	Val	Thr	Met	Ile	Asp
				980					985					990		
35	Glu	Ile	Gly	Ala	Leu	Leu	Asn	Tyr	Ser	Ala	Ser	Thr	Pro	Val	Gly	Thr
			995				1000						1005			
	Pro	Ala	Val	Leu	Pro	Gly	Ser	Arg	Pro	Ala	Val	Leu	Pro	Asp	Gly	Thr
		1010				1015						1020				
40	Val	Thr	Ser	Ala	Asn	Phe	Ala	Val	His	Trp	Thr	Lys	Pro	Ala	Asp	Thr
	1025				1030						1035					1040
	Val	Tyr	Asn	Thr	Ala	Gly	Thr	Val	Lys	Val	Pro	Gly	Thr	Ala	Thr	Val
				1045					1050					1055		
	Phe	Gly	Lys	Glu	Phe	Lys	Val	Thr	Ala	Thr	Ile	Arg	Val	Gln	Arg	Ser
				1060				1065				1070				
45	Gln	Val	Thr	Ile	Gly	Ser	Ser	Val	Ser	Gly	Asn	Ala	Leu	Arg	Leu	Thr
		1075					1080					1085				
	Gln	Asn	Ile	Pro	Ala	Asp	Lys	Gln	Ser	Asp	Thr	Leu	Asp	Ala	Ile	Lys
		1090				1095					1100					
50	Asp	Gly	Ser	Thr	Thr	Val	Asp	Ala	Asn	Thr	Gly	Gly	Gly	Ala	Asn	Pro
	1105				1110					1115					1120	
	Ser	Ala	Trp	Thr	Asn	Trp	Ala	Tyr	Ser	Lys	Ala	Gly	His	Asn	Thr	Ala
				1125					1130					1135		
	Glu	Ile	Thr	Phe	Glu	Tyr	Ala	Thr	Glu	Gln	Gln	Leu	Gly	Gln	Ile	Val
				1140				1145					1150			
55	Met	Tyr	Phe	Phe	Arg	Asp	Ser	Asn	Ala	Val	Arg	Phe	Pro	Asp	Ala	Gly
		1155					1160					1165				
	Lys	Thr	Lys	Ile	Gln	Ile	Ser	Ala	Asp	Gly	Lys	Asn	Trp	Thr	Asp	Leu
		1170				1175					1180					
60	Ala	Ala	Thr	Glu	Thr	Ile	Ala	Ala	Gln	Glu	Ser	Ser	Asp	Arg	Val	Lys
	1185				1190					1195						1200
	Pro	Tyr	Thr	Tyr	Asp	Phe	Ala	Pro	Val	Gly	Ala	Thr	Phe	Val	Lys	Val
				1205					1210					1215		
	Thr	Val	Thr	Asn	Ala	Asp	Thr	Thr	Thr	Pro	Ser	Gly	Val	Val	Cys	Ala
				1220				1225					1230			

	Gly	Leu	Thr	Glu	Ile	Glu	Leu	Lys	Thr	Ala	Thr	Ser	Lys	Phe	Val	Thr
		1235						1240					1245			
	Asn	Thr	Ser	Ala	Ala	Leu	Ser	Ser	Leu	Thr	Val	Asn	Gly	Thr	Lys	Val
		1250					1255					1260				
5	Ser	Asp	Ser	Val	Leu	Ala	Ala	Gly	Ser	Tyr	Asn	Thr	Pro	Ala	Ile	Ile
		1265				1270				1275					1280	
	Ala	Asp	Val	Lys	Ala	Glu	Gly	Glu	Gly	Asn	Ala	Ser	Val	Thr	Val	Leu
					1285					1290					1295	
10	Pro	Ala	His	Asp	Asn	Val	Ile	Arg	Val	Ile	Thr	Glu	Ser	Glu	Asp	His
			1300						1305					1310		
	Val	Thr	Arg	Lys	Thr	Phe	Thr	Ile	Asn	Leu	Gly	Thr	Glu	Gln	Glu	Phe
			1315					1320					1325			
	Pro	Ala	Asp	Ser	Asp	Glu	Arg	Asp	Tyr	Pro	Ala	Ala	Asp	Met	Thr	Val
		1330				1335					1340					
15	Thr	Val	Gly	Ser	Glu	Gln	Thr	Ser	Gly	Thr	Ala	Thr	Glu	Gly	Pro	Lys
		1345				1350					1355				1360	
	Lys	Phe	Ala	Val	Asp	Gly	Asn	Thr	Ser	Thr	Tyr	Trp	His	Ser	Asn	Trp
				1365					1370						1375	
20	Thr	Pro	Thr	Thr	Val	Asn	Asp	Leu	Trp	Ile	Ala	Phe	Glu	Leu	Gln	Lys
			1380						1385					1390		
	Pro	Thr	Lys	Leu	Asp	Ala	Leu	Arg	Tyr	Leu	Pro	Arg	Pro	Ala	Gly	Ser
		1395					1400					1405				
	Lys	Asn	Gly	Ser	Val	Thr	Glu	Tyr	Lys	Val	Gln	Val	Ser	Asp	Asp	Gly
		1410				1415					1420					
25	Thr	Asn	Trp	Thr	Asp	Ala	Gly	Ser	Gly	Thr	Trp	Thr	Thr	Asp	Tyr	Gly
		1425				1430				1435					1440	
	Trp	Lys	Leu	Ala	Glu	Phe	Asn	Gln	Pro	Val	Thr	Thr	Lys	His	Val	Arg
				1445					1450					1455		
30	Leu	Lys	Ala	Val	His	Thr	Tyr	Ala	Asp	Ser	Gly	Asn	Asp	Lys	Phe	Met
			1460					1465					1470			
	Ser	Ala	Ser	Glu	Ile	Arg	Leu	Arg	Lys	Ala	Val	Asp	Thr	Thr	Asp	Ile
		1475					1480					1485				
	Ser	Gly	Ala	Thr	Val	Thr	Val	Pro	Ala	Lys	Leu	Thr	Val	Asp	Arg	Val
		1490				1495					1500					
35	Asp	Ala	Asp	His	Pro	Ala	Thr	Phe	Ala	Thr	Lys	Asp	Val	Thr	Val	Thr
		1505				1510				1515					1520	
	Leu	Gly	Asp	Ala	Thr	Leu	Arg	Tyr	Gly	Val	Asp	Tyr	Leu	Leu	Asp	Tyr
				1525					1530					1535		
40	Ala	Gly	Asn	Thr	Ala	Val	Gly	Lys	Ala	Thr	Val	Thr	Val	Arg	Gly	Ile
			1540						1545					1550		
	Asp	Lys	Tyr	Ser	Gly	Thr	Val	Ala	Lys	Thr	Phe	Thr	Ile	Glu	Leu	Lys
		1555					1560					1565				
	Asn	Ala	Pro	Ala	Pro	Glu	Pro	Thr	Leu	Thr	Ser	Val	Ser	Val	Lys	Thr
		1570				1575					1580					
45	Lys	Pro	Ser	Lys	Leu	Thr	Tyr	Val	Val	Gly	Asp	Ala	Phe	Asp	Pro	Ala
		1585				1590				1595					1600	
	Gly	Leu	Val	Leu	Gln	His	Asp	Arg	Gln	Ala	Asp	Arg	Pro	Pro	Gln	Pro
				1605					1610					1615		
50	Leu	Val	Gly	Glu	Gln	Ala	Asp	Glu	Arg	Gly	Leu	Thr	Cys	Gly	Thr	Arg
			1620						1625					1630		
	Cys	Asp	Arg	Val	Glu	Gln	Leu	Arg	Lys	His	Glu	Asn	Arg	Glu	Ala	His
		1635					1640					1645				
	Arg	Thr	Gly	Leu	Asp	His	Leu	Glu	Phe	Val	Gly	Ala	Ala	Asp	Gly	Ala
		1650				1655				1660						
55	Val	Gly	Glu	Gln	Ala	Thr	Phe	Lys	Val	His	Val	His	Ala	Asp	Gln	Gly
		1665				1670				1675					1680	
	Asp	Gly	Arg	His	Asp	Asp	Ala	Asp	Glu	Arg	Asp	Ile	Asp	Pro	His	Val
				1685					1690					1695		
	Pro	Val	Asp	His	Ala	Val	Gly	Glu	Leu	Ala	Arg	Ala	Ala	Cys	His	His
60			1700						1705					1710		
	Val	Ile	Gly	Leu	Arg	Val	Asp	Thr	His	Arg	Leu	Lys	Ala	Ser	Gly	Phe
		1715					1720						1725			
	Gln	Ile	Pro	Ala	Asp	Asp	Met	Ala	Glu	Ile	Asp	Arg	Ile	Thr	Gly	Phe
		1730					1735					1740				

His Arg Phe Glu Arg His Val Gly